Chlorophyll analysis protocol

1. Turn off lights and put samples on ice.
2. Using tweezers place filter into a 25mL scintillation vial with 10mL of 90% acetone solution, making sure the cap is tight.
3. Record sample name and volume of acetone added.
4. Vortex the filter and acetone solution, place back in box.
5. Rinse tweezers with ethanol and wipe them before touching another filter and repeat.
6. Cover scintillation vials with aluminum foil and store the vials in -20C freezer for 24 hours +/- 4 hours to extract the chlorophyll.

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1. Let samples sit at room temperatures in the dark for at least 10 minutes.
2. Blank the fluorometer using 90% acetone solution in 13mm test tube.
3. Vortex sample and wait for filter debris (if any) to settle for ~30 seconds.
4. Rinse and fill a test tube ¾ of the way and measure raw fluorescence (Rb).
5. After getting Rb, add 3 drops of 10% HCL to the sample, allow signal to stabilize and record reading (Ra).